

Effects of Different Processing Methods on the Iron, Zinc and Other Nutrients Content of Biofortified Beans (*Phaseolus vulgaris L.*)

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Abstract Iron deficiency anemia remains a real public health problem among young children in Cameroon. To reduce it, iron biofortification of legumes was developed to improve the iron nutritional status of the children. This study aimed to assess the effect of different processing methods on the iron, zinc and other nutrients content of biofortified bean cultivars (*Phaseolus vulgaris L.*) from Cameroon. Firstly, a survey was done in the city of Douala on different processes applied to the bean seeds before cooking. The raw and cooked sample of seeds were analysed for proximate and antinutrients composition, using AOAC methods for macronutrients content and atomic absorption spectrophotometry for minerals. The traditional bean cultivar served as control. From the results of the survey, beans treatment were divided into four groups: raw, boiled, soaked and boiled, boiled with limestone. The results showed that different processing methods led to an increase in protein (19.53%-27.66%), and crude fibre (4.46%-7.99%) but a decrease in carbohydrate (62.23%-53.28 %), lipids (5.48%-3.32%) contents. The biofortified bean showed statistically significant differences in iron and zinc contents compared to the traditional bean. Soaked and boiled biofortified bean had higher mineral contents compared to the traditional bean. Processing improved significantly ($p < 0.05$) the nutritional value of the beans by reducing the antinutrient contents. Boiling with limestone was found to have the highest level of reduction effect on the tannin, oxalate, phytate and saponin. Biofortified bean cultivar could be used as food formulation material for infants and young children to prevent micronutrients deficiencies.

Keywords: biofortification, *Phaseolus vulgaris L.*, nutrients, cooking methods, iron

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1. Introduction

Micronutrients malnutrition especially iron deficiency anemia affects at least half of the world's population. The World Health Organization (WHO) reported that an estimated 42% of children aged under 59 months are anemic worldwide. The burden is even higher in Africa, reaching 62.3% [1]. In Cameroon by considering iron deficiency anemia, the most affected age groups are children below 59 months (57%) and women of reproductive age (40%) [2]. This is caused by low intake of daily required amount of micronutrients [3]. However, many authors demonstrated that food fortification used as one of the approaches to overcome micronutrient deficiencies is hard for vulnerable and poor people unable to access fortified

processed food. Therefore, biofortification of popular food crops like beans was proposed as a better intervention to address micronutrient deficiencies [4,5]. It is a process through which the nutritional value, minerals or vitamins levels of a food crop are enhanced through conventional breeding.

Beans and meat are both important sources of iron and approximately 1-7% of this mineral in dietary sources are absorbed when consumed alone [6]. However, according to Barker [7] and Katharine [8], the consumption of a high quantity of meat increases the risk of cardiovascular diseases and some types of cancer because of the presence of other component. In response, intensive efforts were made to find alternative sources of protein from the underutilized leguminous plants in nutrition and in the formulation of new food products.

Common beans are the most grown and largely consumed legume and are affordable sources of proteins in

Cameroon, making them a good choice for biofortification. Beans are one of the main sources of protein, plant-derived micronutrients, and minerals. Common beans (*Phaseolus vulgaris* L.) are rich in protein (14-24%) and micronutrients (Fe = 7-8 mg/100g and Zn = 2.5 3.5 mg/100g) [9]. However, the health benefits of beans are associated with their processing methods. Beans should be cooked or processed before intake. These processes increase the bioavailability of nutrients and reduces flatulence (raffinose oligosaccharides) and antinutrients factors [6]. According to many authors, the health benefits of beans are associated with their processing methods. Beans should be cooked or processed before intake [10,11].

Concerning different processing techniques, the studies conducted in Yaounde [12] showed that simple boiling, boiling with sodium bicarbonate, soaking and boiling, boiling with *Echinops giganteus* bark powder reduce significantly ($P < 0.05$) the level of antinutrients, thus improve protein and starch digestibility of bean. Moreover, according to Ranilla et al. [13], cooking imparts desirable sensory properties to grains. Much information is available in the literature on nutrient content and potential contribution of different varieties of raw beans [9,14], but information on biofortified bean is scanty. This study aims to evaluate the effect of different processing methods commonly used by households on the proximate, mineral and antinutrients composition of biofortified bean cultivars (*Phaseolus vulgaris* L.).

2. Materials and Methods

2.1. Sample Collection and Preparation

This was a descriptive study. The study have begun with a survey of bean consumers (traders, households) without distinction of sex in the city of Douala. The purpose of this survey was to identify the different processing methods of beans commonly used in households. The survey took place from October 2020 to December 2020 in Douala (Cameroon). At the end of the survey, three (03) cooking methods most commonly used by householders were identified. Iron biofortified bean variety (FEB 192) and traditional bean variety (PH 201) were purchased from Institute of Agricultural Research for Development (IRAD) of Foumbot station (West region of Cameroon). The two varieties were selected due to consumer appreciation for traditional variety (PH 201) and the availability of iron biofortified bean for FEB 192 (Figure 1).



(a): FEB 192 (large red seed); (b): PH 201 (small red seed).

Figure 1. Picture of the two bean cultivars (*Phaseolus vulgaris* L.)

The two varieties of beans FEB 192 and PH 201 were transported to the laboratory where they were sorted by removing dirt or broken beans. Then, they were washed thoroughly, to remove soil and all foreign particles. The seeds obtained were divided into 4 batches. The first batch was used for raw sample analysis, the second batch was boiled, the third batch was soaked (12h) and boiled and the fourth batch was boiled with 15g of limestone. For the first batch, 500g of each raw seeds were finely ground to a fine powder with a Kenwood blender after washing and drying to a constant weight at 50°C for 72 hours. The resultant flour was packaged in an airtight vessel in readiness for analysis. The vessel was appropriately labelled. For the second batch, 500g of bean seeds of each variety were boiled at 100°C for 1h30mn in 2.5L of distilled water in the proportion of 1:5 (w/v). The cooked seeds were dried to a constant weight at 50°C for 72 hours with frequent turning and mill. The resultant flour was packaged in an airtight appropriately labelled vessel in readiness for analysis. For the third batch, 500g of bean seeds of each variety were firstly soaked in distilled water in a 1:5 proportion (w/v) for 12 hours at room temperature 28°C. Subsequently, the soaking water was not removed. Secondly, the soaked bean seeds were boiled at 100°C for 1h30mn in 2.5L of distilled water in the proportion of 1:5 (w/v). The cooked seeds were dried to a constant weight at 50°C for 72 hours with frequent turning and mill. The resultant flour was packaged in an airtight appropriately labelled vessel in readiness for analysis. For the fourth batch, 500g of bean seeds of each variety were boiled at 100°C during the boiling, 15g of limestone were added and the all were left boiling 45mn of distilled water in the proportion of 1:5 (w/v). The cooked seeds were dried to a constant weight at 50°C for 72 hours with frequent turning. The resultant flour was packaged in an airtight appropriately labelled vessel in readiness for analysis.

2.2. Analysis

Moisture, proteins, lipid, crude fibres, ashes and carbohydrates content were determined using standard methods of Association of Official Analytical Chemists [15]. The samples were analyzed in triplicate.

2.3. Proximate Composition

The moisture was determined in an oven set at 105°C, according to standard procedures detailed by AOAC [15] during 72h (to the constant weight). The total nitrogen contents were determined by the Kjeldahl method, as described by AOAC, and the protein content was calculated by multiplying result by 6,25. Lipids content were evaluated by Soxhlet extraction according to the method described by AOAC [15], using hexane as the extractor. The ashes content were determined by calcination in a furnace at 550°C. The total fiber content was determined gravimetrically after delipidation of bean powder using the method described by AOAC [15].

2.4. Mineral Analysis

The mineral contents (iron, zinc, magnesium, phosphorus and calcium) of different samples were determined by

AOAC method N°968.08 [16]. About 100 g of powder for each raw and cooked bean seeds cultivars were oven dried at 105°C for 72 hours. After drying, 5g of beans were separately weighed into crucibles and maintained at 550°C for 24 hr. The ashes were cooled in desiccators and then weighed. After weighing, the ashes were dissolved in a solution of 1:1 ratio of H₂O: HCl, in which the concentration of the final mixture was 6N HCl. The contents of iron, zinc, magnesium, phosphorus and calcium were determined by atomic absorption spectrophotometer (Shimadzu UNICAM 919, England), while total phosphorus concentration was measured by colorimetric spectrophotometer after incubation with Molybdo-vanadate solution. Potassium and sodium levels were determined by digesting the ashes of the samples with perchloric acid and nitric acid, and then taking the readings on Jenway digital flame photometer/spectronic 20 [15].

2.5. Antinutrients Analysis

Tannins content were determined using ferric reagent in an acidic alcoholic medium using gallic acid as standard [17]. The absorbance was readed at 550 nm. The total oxalate content was assessed by titration with KMnO₄ after acid digestion [18]. The phytate content was determined by titration with iron III solution after acid digestion [19]. Saponin contents were determined by weight difference after extraction in solvent as described by Obadoni and Ochuko [20].

2.6. Statistical Analysis

Data were expressed as mean ± standard deviation of triplicate samples and measurements. Analysis of variance (ANOVA) and the comparison of means (Tukey's test,

P<0.05) were applied using IBM/SPSS 20.0 software (Statistical Package of Social Science) for Windows. The sodium to potassium (Na/K) and calcium to phosphorus (Ca/P) ratio were calculated in the samples. The energy value (E) per 100g of bean cultivars was obtained using Atwater [21] conversion [21] factors as follows:

$$E(\text{Kcal}) = \text{Proteins}(\%) \times 4 + \text{carbohydrate}(\%) \times 4 + \text{lipids}(\%) \times 9.$$

Determination of molar ratio of antinutrients to minerals was predicted by dividing the mole of antinutrient by the mole of minerals [22].

3. Results

The results obtained in the study are summarised in Table 1 - Table 4 and comprising macronutrients contents and the energy value, mineral contents, antinutrient contents and the molar ratio of antinutrients to mineral of biofortified bean seeds according to the various treatments.

Table 1 shows the moisture, proteins, lipids, carbohydrates, crude fibre, ashes contents of biofortified and traditional beans according to different treatments. The two cultivars of beans were high in proteins and carbohydrates, moderate in ashes and crude fibres but low in fats and moisture contents after all the treatments. The proteins and carbohydrates content of biofortified beans were higher (P<0.05) than that of traditional beans. It appeared that the soaked and boiled seeds had the highest protein contents (29.28±4.87g/100g DW; 27.66±1.63g/100g DW respectively for the PH 201 and FEB 192). The soaked and boiled seeds had the highest energy value 356.79 Kcal/100g DW (PH 201) and 355 Kcal/100g DW (FEB 192).

Table 1. Proximate composition and energy of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.)

Cultivars Parameter (%)	PH 201				FEB 192			
	Raw	Boiled	SoB	BSb	Raw	Boiled	SoB	BSb
Moisture	8.17±0.56 ^a	8.26±1.35 ^a	8.61±0.53 ^a	8.31±0.60 ^a	3.97±2.02 ^a	5.88±2.59 ^{ab}	4.55±0.51 ^a	7.87±0.60 ^b
Protein	18.72±0.82 ^a	19.05±0.43 ^a	29.28±4.87 ^c	24.41±1.63 ^c	19.53±0.00 ^a	26.04±0.00 ^b	27.66±1.63 ^b	21.96±0.82 ^d
Lipids	5.04±1.05 ^a	3.09±0.80 ^{ab}	2.51±1.05 ^b	3.44±0.69 ^{ab}	5.48±1.66 ^a	3.57±0.39 ^{ab}	3.32±0.66 ^b	4.79±1.37 ^{ab}
Carbohydrates	57.79±1.81 ^a	59.62±0.04 ^a	49.38±2.13 ^{ac}	55.63±4.18 ^c	62.23±1.94 ^a	53.28±0.35 ^b	53.79±2.19 ^b	55.68±0.42 ^b
Ashes	3.23±0.02 ^a	3.68±0.14 ^b	2.57±0.24 ^c	3.09±0.04 ^a	4.33±0.13 ^a	3.24±0.17 ^b	3.09±0.03 ^b	3.08±0.02 ^b
Crude fibre	7.05±0.07 ^a	6.30±0.19 ^b	7.65±0.19 ^c	5.12±0.09 ^d	4.46±0.13 ^a	7.99±0.05 ^b	7.59±0.11 ^c	6.62±0.25 ^d
Energy (Kcal)	351.40	342.49	337.23	351.12	376.36	349.41	355.68	353.67

Values are means ± SD of triplicate determinations, Means within the same line (for the same cultivar) with different superscripts significantly different at p<0.05. SoB = Soaked and boiled; BSb = Boiled with limestone.

Table 2. Mineral contents of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.)

Cultivars Mineral (mg/100g DW)	PH 201				FEB 192			
	Raw	Boiled	SoB	BSb	Raw	Boiled	SoB	BSb
Fe	7.75±0.64 ^a	8.72±0.64 ^b	8.45±1.71 ^b	4.93±0.00 ^d	8.73±0.64 ^a	8.97±1.28 ^{ab}	7.28±1.50 ^c	5.46±0.86 ^d
Zn	2.23±0.00 ^a	2.73±0.00 ^b	2.64±0.00 ^c	1.90±0.00 ^d	3.10±0.00 ^a	3.38±0.00 ^b	3.70±0.00 ^c	3.14±0.00 ^d
Na	2.81±0.02 ^a	2.76±0.04 ^b	2.32±0.01 ^c	27.26±0.00 ^d	2.83±0.00 ^a	1.88±0.02 ^b	1.86±0.00 ^b	42.29±0.04 ^d
Ca	63.12±0.08 ^a	70.8±0.08 ^b	93.96±0.04 ^c	97.26±0.02 ^d	86.20±0.02 ^a	93.14±0.11 ^b	86.84±0.04 ^c	106.89±0.04 ^c
Mg	242.85±0.06 ^a	197.4±0.04 ^b	179.25±0.08 ^c	188.7±0.02 ^d	234.9±0.04 ^b	187.5±0.00 ^b	168.15±0.03 ^c	192.3±1.80 ^d
P	67.23±5.25 ^a	71.43±2.10 ^a	58.30±11.03 ^a	96.11±7.90 ^d	67.33±4.05 ^a	63.02±13.65 ^a	57.77±12.60 ^a	59.87±15.76 ^a
K	20.69±0.05 ^a	22.25±0.01 ^b	20.24±0.00 ^c	19.10±0.08 ^d	23.33±0.00 ^a	28.80±0.02 ^b	22.86±0.00 ^c	25.90±0.04 ^d
Na/K	0.14	0.12	0.11	1.43	0.12	0.06	0.08	1.63
Ca/P	0.94	0.99	1.61	1.01	1.28	1.48	1.50	1.79

Values are means ± SD of triplicate determinations, Means within the same line (for the same cultivar) with different superscripts significantly different at p<0.05. SoB = Soaked and boiled; BSb = Boiled with limestone. Na/K= sodium to potassium ratio; Ca/P = calcium to phosphorus ratio.

Table 3. Antinutrient contents of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.)

Cultivars Parameter (mg/100g DW)	PH 201				FEB 192			
	Raw	Boiling	SoB	BSb	Raw	Boiling	SoB	BSb
Phytates	0.15±0.09 ^a	0.10±0.00 ^b	0.06±0.00 ^c	0.06±0.01 ^b	1.89±0.00 ^a	0.06±0.00 ^a	0.08±0.01 ^a	0.04±0.01 ^a
Tannins	0.61±0.06 ^a	0.02±0.00 ^b	0.09±0.00 ^c	0.09±0.01 ^c	0.28±0.03 ^a	0.07±0.01 ^b	0.24±0.01 ^c	0.16±0.01 ^d
Oxalates	0.70±0.04 ^a	0.31±0.00 ^b	0.22±0.01 ^c	0.31±0.03 ^b	0.58±0.04 ^a	0.25±0.02 ^b	0.31±0.03 ^{bc}	0.40±0.08 ^c
Saponins	5.10±0.05 ^a	4.83±0.23 ^a	4.65±0.15 ^a	4.63±0.48 ^a	5.33±0.18 ^a	4.85±0.00 ^b	4.40±0.00 ^c	4.40±0.10 ^c

Values are means ± SD of triplicate determinations, Means within the same line (for the same cultivar) with different superscripts significantly different at $p < 0.05$. SoB = Soaked and boiled; BSb = Boiled with limestone.

Table 4. Calculated molar ratios of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.)

Cultivars Parameters	PH 201				FEB 192				Standard value
	Raw	Boiling	SoB	BSb	Raw	Boiling	SoB	BSb	
(Phytates/Ca) ^a	1.443x10 ⁻⁴	8.58x 10 ⁻⁵	3.97x10 ⁻⁵	3.74x10 ⁻⁵	1.33x10 ⁻³	3.91x10 ⁻⁵	5.59x10 ⁻⁵	2.27x10 ⁻⁵	< 0.24
(Phytates/Fe) ^b	1.64x10 ⁻³	9.70x10 ⁻⁴	6.01x10 ⁻⁴	1.01x10 ⁻³	1.83x 10 ⁻²	5.66x10 ⁻⁴	9.30x10 ⁻⁴	6.12x10 ⁻⁴	< 0.15
(Phytates/Zn) ^c	6.66x10 ⁻³	3.63x10 ⁻³	2.25x10 ⁻³	3.13x10 ⁻³	6.04x10 ⁻²	1.76x10 ⁻³	2.14x10 ⁻³	1.26x10 ⁻³	< 10
(Oxalates/Ca) ^d	3.47x10 ⁻³	1.37x10 ⁻³	7.33x10 ⁻⁴	9.97x10 ⁻⁴	2.10x10 ⁻³	8.40x10 ⁻⁴	1.12x10 ⁻³	1.17x10 ⁻³	< 1
(Phytates*Ca/Zn) ^e	1.05x10 ⁻²	6.41x10 ⁻³	5.28x10 ⁻³	7.59x10 ⁻³	1.30x10 ⁻¹	4.09x10 ⁻³	4.64x10 ⁻³	3.37x10 ⁻³	0.5

SoB = Soaked and boiled; BSb = Boiled with limestone; ^amg of phytate/molecular weight of phytate: mg of calcium/molecular weight of calcium. ^bmg of phytate/molecular weight of phytate: mg of iron/molecular weight of iron. ^cmg of phytate/molecular weight of phytate: mg of zinc/molecular weight of zinc. ^dmg of oxalate/molecular weight of oxalate: mg of calcium/molecular weight of calcium. ^e(mg of calcium/molecular weight of calcium) (mg of phytate/molecular weight of phytate)/(mg of zinc/molecular weight of zinc).

The mineral contents of biofortified and traditional beans seeds (*Phaseolus vulgaris* L.) according to the different treatments, expressed in mg/100g dry weight (DW), are shown in Table 2. The iron contents of biofortified bean were higher 8.73 (raw) than 7.75 (raw) of traditional bean seeds ($P < 0.05$). The two varieties were high in magnesium, iron and potassium but low in zinc ($P < 0.05$). The Ca/P values ranged from 1.28 to 1.79 for the biofortified bean cultivar and from 0.94 to 1.61 for the traditional bean. The Na/K values were in the range of 0.06 to 1.63 for the biofortified variety. The control sample had the highest value of Na/K (1.43) in sample boiled with limestone while the least value (0.11) was held by sample soaked for 12h and boiled.

Table 3 shows the antinutrient contents of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.) expressed in mg/100g DW. The two cultivars (raw) had the same rate of saponins. All processes led to significant reduction in the levels of all the antinutrients ($P < 0.05$).

Table 4 shows the calculated molar ratios of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.). The molar ratios of phytate to calcium (Phy/ca) ranged from 3.74x10⁻⁵ to 1.443x10⁻⁴ for the traditional bean and 2.27x10⁻⁵ to 1.33x10⁻³ for the biofortified beans. The ratios of phytate to iron (Phy/Fe) of the two cultivars varied from 5.66x10⁻⁴ to 1.83x 10⁻².

4. Discussion

In order to improve the nutritional quality of beans, many cooking methods (such as boiling, soaking, boiling with limestone, boiling with sodium bicarbonate, soaking with sodium bicarbonate) are commonly used by people for preparing beans [12]. All of these processes reduce antinutritional factors but other phenomena may also occur, such as losses of the macronutrients and micronutrients, particularly minerals, during the processes of soaking and cooking [6].

All the bean samples are low in moisture and fell within the recommended range of 0-13.5% [24]. The value of 8.17% (traditional raw seeds) is higher than 3.97 % of biofortified beans (raw). The 3.97% moisture content of the biofortified bean obtained for the raw seed was low when compared to those values obtained and reported by Alayande et al. [25] for brown (5.08) and white (3.56) beans and the one reported by Brigide et al. [6] for Porto real (15.24) and Pirata (13.89) biofortified bean cultivars. It can be seen that all processes (boiling, roasting and boiling, boiling with limestone) increases the water content. This phenomenon can be explained by the fact that during these treatments the cells absorb water. The low moisture content of PH 201 and FEB 192 remains an asset in storage and preservation of the nutrients.

The proteins content ranged from 19.53% -27.66% for biofortified beans and 18.72%-29.28% for the traditional beans. The values lay within the range (18-23%) recorded by Faldu et al. [26] for raw and processed kidney bean seeds. The flour sample from the seed of biofortified bean soaked for 12h had the highest value (27.66%) while the traditional sample boiled with limestone had the highest value (29.28%). A significant difference ($P < 0.05$) is observed between the protein contents of the raw and the processed seeds. The boiled seeds (26%), soaked and boiled (27.66%) and boiled with limestone (21.96%) had the highest content compared to the raw seeds (19.53%). The different processing methods enhanced crude protein content of the traditional and biofortified bean. This could be justified by changes in the association and dissociation properties of proteins caused by heating. During cooking, there is disintegration of the crude protein into amino acids and therefore the heat treatment induces changes in the structure of the proteins, which can inactivate the antinutrients, thus increasing the digestibility and the biological values of the protein of the bean [27]. This observation is in agreement with the findings of Audu et al. [28] for sample processed by roasted, fermented and boiled. We observed an increase in nutrients particularly protein. Processed biofortified bean cultivars can be a

good source of protein for the formulation of infants foods.

The lipid content were in the range of 3.32-5.48% for FEB 192 and 2.51-5.04% for PH 201. All the samples (raw and processed) biofortified and traditional bean cultivars contained low fat. Lipid content of the biofortified and traditional bean cultivars obtained for raw and processed seed was high when compared those values obtained and reported by Lajide et al. [29] in Ghana and by Farinde et al. [24] for Lima bean flour (1.14%) after cooking. However, the different processing methods reduced fat content of our two bean cultivars. This could be due to loss of total solids during soaking prior to further processing and denaturation of the fat by heat processing and leaching into the processing water [30]. Boiling with limestone resulted in increase the lipid content of the cultivars. This could be explained by the fact that limestone breaks the cells and membrane during boiling and then, lipids are liberated hence making lipids available [30]. Beans are not a good source of lipid. Low lipid content in the beans is an advantage, as this will reduce the risk of heart attack and increased blood cholesterol level [31].

The crude fibre contents in the two cultivars ranged between 4.46% in the raw biofortified bean to 7.99% in the boiled biofortified bean. Processing also increased the crude fibre in the all batches for biofortified beans. However, crude fibre content decreased in boiling (6.30%) and boiling with limestone (5.12%) of traditional beans. The crude fibre content of the biofortified bean flours was 4.46-7.99%. The traditional sample had the highest value (7.65%) while the least value (5.12%) was held by the sample soaked for 12h and boiling. The values were within the range of 4.5%-17.5% crude fibre content for different bean cultivars in America as reported by Messina et al. [32]. The range of values was less than 17.41%-28.20% reported by ENV. [33] for 13 bean cultivars in Paris. The high rate of crude fibre in the processed sample could be explained by the fact that heat treatments can have variable effects on crude fibre and that cooking causes disruption of the cellular components of beans (cellulose, hemicellulose, lignin, pectin and gums). The cooking process results in interactions between proteins and lipids and it leads to qualitative and quantitative changes in the composition of total fibre of cooked foods compared to that of raw foods [6]. Our findings suggest that the value of crude fibre is within acceptable limit which helps to maintain the health of gastrointestinal tract [34].

The ash content of the biofortified bean cultivars was 3.08- 4.33%. The ash content of traditional bean cultivars was 2.57%-3.68% (Table 1). The values were within the range of 3.94-4.82% ash content for Lima bean (*Phaseolus Lunatus*) flour reported by Oraka et al. [35] and Duru et al. [36] for Velvet beans (3.18%-3.44%). There was a reduction in the ash content in the processed seeds except for the boiling traditional bean cultivar (3.68%). Low ash content in the bean could be due to leaching of salts and minerals into the cooking water [24].

The carbohydrates contents of our study suggest that biofortified bean cultivar could be a good supplement to which could be use for feed formulation. The carbohydrate values were low compare to those reported

by Alayande et al. [25] for white and brown beans in Jos North (Nigeria) (56.80%-60.47%) and (55%-65%), (82.90%-87.29%) reported for the common beans (*Phaseolus vulgaris*) respectively by Alayande et al. [25] and Idoko et al. [37]. These values are close (51% to 41%) to those reported for the processed Velvet beans (*Mucuna Pruriens*) [36]. All the processing methods decrease the carbohydrate contents except for the boiling traditional seeds (59.62%). Similar observations were obtained on the seeds in Nigeria [31]. According to fernandes et al. [27], that reduction of the amounts of available carbohydrates could be due by the fact that carbohydrates being the water soluble compounds and then, they have been hydrolyzed and diffused in the soaking and cooking water. This low carbohydrate content in our study suggests that biofortified bean cultivar could be a potential good source of formulation for people suffering from diabete.

The energy content of our seeds depends on some of the products of each major food (proteins lipids and carbohydrates). The soaking and boiled biofortified bean cultivar had the highest value (355.68), followed by boiled with limestone (353.67) and raw seeds (376.36). The energy value of biofortified bean in this study showed that the biofortified bean cultivar has an energy concentration more favourable than extruded biofortified bean flours [38].

The result of mineral contents of raw and processed biofortified and traditional beans cultivars (*Phaseolus vulgaris* L.) shown in Table 2 indicate that the raw seeds of the two cultivars and the processed seeds contained appreciable amount of minerals (magnesium, iron, calcium, phosphorus and zinc). The most abundant mineral in the raw seed samples was magnesium (242.85 mg/100g and 234.9 mg/100g), calcium (63.12 mg/100g and 86.20 mg/100g) respectively for the traditional and biofortified bean cultivars. The raw seeds of biofortified beans were also rich in iron (8.73 mg/100g), phosphorus (67.33 mg/100g) and potassium (23.33 mg/100g). Processing significantly ($P < 0.05$) decreased the mineral content (except for the calcium) in the raw seeds of the two cultivars. Reduction of the mineral content of the processed samples could be due to leaching of the minerals into the cooking water during boiling. However, according to Bamigboye et al. [31], increase of calcium content could be explained by the fact that during cooking the bonds between antinutrients and calcium were broken, which led to release of calcium into the cooking water resulting a higher concentration [31]. The results obtained for the two cultivars in magnesium are agreement with the report of the literature [24,39]. Furthermore, contradicts the reports of Mananga et al. [9] that potassium is the most abundant mineral in ten red bean cultivars. The value for sodium in the raw samples is lower than values reported by Hassan and El Syiad. [40] for white bean seeds.

Several authors found iron levels in 100g varied from 7.4 mg to 11.5 mg [28]. Iron is required for oxygen to travel to tissues and organs. It helps to carry oxygen throughout the body in form of heamoglobin and myoglobin, it is an integral part of many proteins and enzymes and it also helps in energy metabolism [41]. Our findings are within the range of RDA for iron (7-16 mg/100g) [42]. Biofortified bean cultivars was good

source of iron and can be used for formulation for infants with iron deficiency anemia. Magnesium is an activator of many enzymes system and maintains the electrical potential in nerves [43]. It values in this study are above the recommended daily magnesium requirements (80-130mg for children) [43].

The very low sodium (except for boiling with limestone samples) and high potassium content of the raw bean cultivars is advantageous and make them suitable for management of metabolic syndrome. Indeed, treatment of high blood pressure requires a low sodium and high potassium for intra and extracellular electrolyte balance [31]. Potassium is nutritionally important for pH regulation and the proper functioning of carbohydrate and protein metabolism [44]. For this reason, bean seeds are an excellent food to cover daily potassium requirements (800 to 1600 mg/100g) for children of 2 to 9 years old [45]. The Na/K ratio in the body is of great concern for the prevention of high blood pressure. The Na/K ratio less than one is required. So, biofortified bean seeds could be used in the prevention of high blood pressure.

The value for zinc in the raw flour samples is higher than values reported by Faldu et al. [26] for chickpea (*Cicer arietinum*) and kidney bean (*Phaseolus vulgaris*). Zinc boosts the health of our hairs, plays a role in the proper functioning of some sense organs such as ability to taste and smell [6]. The zinc contents of our samples are lower than the RDA of 3 mg/day (children) and 11 - 12 mg/day (pregnant women) [43]. FEB 192 are an excellent food to cover daily zinc requirements.

Calcium is known as a macroelement necessary for the development of teeth, bones and the release of hormones [46]. Daily calcium requirements are 700-1300 mg for children [47]. Biofortified bean cultivars should be prepared and consumed with calcium rich foods for a good nutritional balance. The value for phosphorus in the raw flour samples is lower than values reported by Otitoju et al. [48] for the Cowpea (*Vigna Unguiculata*). Phosphorus enables the fixation of calcium in the bones by decreasing its urinary excretion and takes part in the mechanism of energy storage and release [30]. According to Audu and Aremu. [28], phosphorus is always found with calcium in the body both contributing to the blood formation and supportive structure of the body. This led to the concept of calcium phosphorus ratio. If the Ca/P ratio is low, calcium will be low and there will be high phosphorus intake which leads to calcium loss in the urine more than normal. If the Ca/P ratio of any food is above one that food is considered "good" and "poor" if the ratio is less than 0.5, while a Ca/P ratio above two helps to increase the absorption of calcium in the small intestine. The Ca/P ratio in the biofortified bean cultivars particularly processed samples were above one. This suggests that biofortified bean could be considered good food.

Variations in the micronutrient contents of cultivars can be attributed to a number of factors: plant characteristics, such as plant age, maturity, species, variety, cultivar, environmental features, such as climate, soil, rainfall, season and processing factors, such as storage time, temperature, method of preservation, and preparation of food.

Antinutrients are found at some level in almost all foods. However, their levels are reduced through various traditional methods [49]. The findings of this study

revealed that all the processing methods significantly ($p < 0.05$) reduced antinutrients content in PH 201 and FEB 192 bean seeds. The amount of oxalates ranged from 0.70% (raw) to 0.22% (soaked and boiled) in traditional bean and 0.58% (raw) to 0.25% (boiled) in biofortified bean. The similar results have been reported by Farinde et al. [24] for processing Lima beans (*Phaseolus Lunatus*). In the literature, it was reported that oxalates reduce the availability of essential nutrients [50]. Diet high in oxalate has been reported to increase the risk of development of kidney stone [51]. The oxalate to calcium (Oxalates/Ca) ratio of the biofortified beans varied from 2.10×10^{-3} to 8.40×10^{-4} (Table 4). This result implies that oxalate cannot have any adverse effects on bioavailability of dietary calcium in these seeds.

Tannin contents ranged between 0.61% (raw) to 0.09% (soaked and boiled and boiled with limestone) and 0.28% (raw) to 0.07% (boiled) in PH 201 and FEB 192 respectively. Processing significantly reduced the tannin contents in traditional and biofortified beans (more than 90%). The low tannin contents in the processed biofortified beans are similar to those values obtained and reported by Silva et al. [52] for the biofortified carioca bean (2.15% to 0.02) and the common bean (0.42 to 0.03%), and one reported by Mugabo et al. [53] for bean flours (0.90%-0.19%). According to Toledo et al. [54] tannins can interact with protein and interfere with digestibility of beans, decreasing the hydrolysis of phaseolin. Soaking and cooking are processing that can influence the amount of tannins.

Concerning to the determination of phytates, the concentration varied from 0.15% (raw) to 0.06 (soaked and boiled and boiled with limestone) in the traditional and 1.89% (raw) to 0.04% (boiled with limestone) in biofortified beans. These values are less than that of processed bean flours in Rwanda (2.42% -1.09%) [53]. According to Fabbri and Crosby. [55], boiling reduces phytates content of vegetables. However, a longer cooking time often results in greater reduction of phytates. Boiling with limestone and simple boiling were found to significantly reduced ($p < 0.05$) all the phytates evaluated in the biofortified bean seeds. The molar ratio of phytate to iron (Phy/Fe) of biofortified bean ranged from 6.12×10^{-4} to 1.83×10^{-2} . According to Fekadu et al. [56], the phytate/iron molar ratios < 0.15 is indicative of good iron bioavailability. The result indicated that the biofortified bean contain the phytate/iron molar ratios of less than the critical value this implies the good availability of iron. The phytate/calcium molar ratio varied from 2.27×10^{-5} to 1.33×10^{-3} (Table 4). The values are below the standard value (< 0.24), this implies good calcium bioavailability of biofortified bean [56]. The molar ratio of phytate to zinc (Phy/Zn) ranged from 6.04×10^{-2} to 1.26×10^{-3} , was less than 10, which is the critical molar ratio of Phy/Zn [56]. This indicates that biofortified bean have adequate availability of zinc.

Saponin contents varied from 5.10% (raw) to 4.63% (boiled with limestone) and 5.33% (raw) to 4.40% (soaked and boiled, boiled with limestone) in traditional and biofortified beans respectively. These values were higher than the value (0.05%) reported for boiling and roasting kidney bean seeds flour in Nigeria [58]. Boiling with limestone were found to significantly reduced ($p < 0.05$) saponins evaluated in the samples. Saponins are known

for their foaming properties in aqueous solution astringent taste and haemolytic activity on red blood cells [14]. Saponins are secondary metabolites with antibacterial and antihelminthic activities [59]. In conclusion, the different processes (boiling with limestone, boiling, soaking and boiling) applied reduce significantly the antinutrient contents and make the minerals bioavailable.

5. Conclusion

Biofortified bean cultivars (FEB 192) were good sources of proteins, carbohydrates, crude fiber, iron, zinc, magnesium and potassium. The processing methods used by households improve the protein contents and reduce significantly the values ($P < 0.05$) of antinutrients (oxalates, phytates, saponines and tannins). However, boiling, soaking and boiling, boiling with limestone decrease mineral contents (except calcium) slightly but remain high enough to cover the recommended daily needs for Fe, Zn, Ca, Mg in children. The processing method have improved the bioavailability and absorption of iron, zinc and calcium. Biofortified bean cultivars may be exploited in food formulation to prevent malnutrition and micronutrient deficiencies.

Conflict of Interests

The authors have not declared any conflict of interests.

References

- [1] Mbunga, B.K., Mapatano, M.A., Strand, T.A., Gjengedal, E.L.F., Akilimali P.Z. and Engebretsen I.M.S. (2021). Prevalence of anemia, iron deficiency anemia, and associated factors among children aged 1-5 years in the Rural, malaria endemic setting of Popokabaka, Democratic Republic of Congo: A cross-sectional study. *Nutrients*, 13, (1010): 2-13.
- [2] EDSC-MICS (Enquête de Démographie et de Santé à Indicateurs multiples) (2018). Rapport préliminaire. Institut national de la statistique. EDSC-MICS, pp 79.
- [3] Kana, M., Mananga, M., Tetanye, E. and Gouado, I. (2015). Risk factors of anemia among young children in rural Cameroon. *International Journal of current Microbiology and Applied Science*, 4(2): 925-935.
- [4] Pfeiffer, W. and McClafferty, B. (2007). Harvest Plus: Breeding crops for better nutrition. *Crop Science*, 47: 88-105.
- [5] Glahn, R., Tako, E., Hart, J., Haas, J., Lung'aho, M. and Beebe, S. (2017). Iron bioavailability studies of the first generation of iron biofortified beans releases in Rwanda. *Nutrients*, 9: 1-11.
- [6] Brigide, P., Canniatti-Brazaca, S. and Silva, MO. (2014). Nutritional characteristics of biofortified common beans. *Food Science and Technology*, 34 (3): 493-500.
- [7] Barker, B. (1996). Wild plants for human nutrition in the Sahelian zone. *Journal of Sand and Environment*, 11: 61-63.
- [8] Katharine, W. (2002). Healing foods Newlanark, MLI9DJ. Scotland.
- [9] Mananga, M.J., Kouandjoua, B.D., Kotue, T.C., Bebe, F., Djuikwo, N.R., Mbassi, M.G., Kuagny, B., Djouhou, M., Fokou, E. and Kana, S.M. (2021). Nutritional and antinutritional characteristics of ten Red bean cultivars (*Phaseolus vulgaris* L.) from Cameroon. *International Journal of Biochemistry Research & Review*, 30 (4): 1-14.
- [10] Huber, K., Brigide, P., Bretas, E.B. and Canniatti-Brazaca, S.G. (2016). Phenolic acid, flavonoids and antioxidant activity of common brown beans (*Phaseolus vulgaris* L.) before and after cooking. *Journal of Nutrition & Food Sciences*, 6 (551): 1-7.
- [11] Mamiro, P.S., Mwanri, A.W., Mongi, R.J., Chivaghula, T.J., Nyagaya, M. and Ntwenya, J. (2017). Effect of cooking on tannin and phytate content in different bean (*Phaseolus vulgaris*) varieties grown in Tanzania. *African journal of biotechnology*, 16(20): 1186-1191.
- [12] Mananga, M.J., Eyili, N.J., Kotue, T.C., Kouandjoua, N.B.D. and Fokou, E. (2022). Effect of different processing methods on the nutritional value of red and white bean cultivars (*Phaseolus vulgaris* L.). *Journal of Food and Nutrition Sciences*, 10(1): 27-35.
- [13] Ranilla, L.G., Genovese, M.I. and Lajolo, F.M. (2009). Effect of different cooking conditions on phenolic compounds and antioxidant capacity of some selected Brazilian bean (*L.*) cultivars. *Journal of Agricultural and Food Chemistry*, 57(13): 5734-5742.
- [14] Kwuimgoin, I., Kotue, T., Kansci, G., Mananga, M. and Fokou, E. (2018). Assessment of mineral contents and antioxidant activities of some bean seeds (*Phaseolus vulgaris* L.) from West Cameroon region. *International Journal of Food and Nutrition Science*, 7 (3): 58-63.
- [15] AOAC (2005). Official Methods of Analysis: Association of Official Analytical Chemists Methods. AOAC. International. Gaithersburg, MD. USA.
- [16] AOAC (1995). Official Methods of Analysis: Association of Official Analytical Chemists Methods. AOAC 16th ed. p13. Washington, DC.
- [17] Ndhlala, A., Kasiyamhuru, A., Mupure, C., Chitindingu, K., Benhura, M. and Muchuweti, M. (2007). Phenolic composition of *Flacourtiandica*, *Opuntia megacantha* and *Sclero cayabirrea*. *Food chemistry*, 103 (1): 82-87.
- [18] Aina, V., Sambo, B., Zakari, A., Haruna, H., Umar, K. and Akinboboye, R. (2012). Determination of nutritional and antinutritional content of *Vitisvinifera* (Grapes) grown in Bomo (Area C) Zaira, Nigeria. *Advanced Journal of Food Sciences and Technology*, 4 (6): 225-228.
- [19] Olayeye, L., Owolabi, B., Adesina, A. and Isiaka, A. (2013). Chemical composition of red and white cocoyam (*Colocasia esculenta*) leaves. *International Journal of Sciences and Research*, 11 (2): 121-5.
- [20] Obadoni, B. and Ochuko, P. (2001). Phytochemical studies and comparative efficacy of the extracts of some hemostatic plants in Edo and Delta States of Nigeria. *Global Journal of Pure and Applied Science*, 8: 203-218.
- [21] Meril, A.L. and Watt, B.K. (1955) Energy Value of Food Basis and Derivation. Handbook N^o 74, US Department of Agriculture, Washington DC.
- [22] Norhaizan, M.E. and Norfaizatul, A.A. (2009). Determination of phytate, iron, zinc, calcium contents and their molar ratios in commonly consumed raw and prepared food in Malaysia. *Malaysian Journal of Nutrition*, 15(2): 213-222.
- [23] Opara, C.I., Egbuonu, C.A. and Obike, C.A. (2017). Assessment of proximate, vitamins, minerals and antinutrients compositions of unprocessed *Vigna aconitifolia* (Moth Bean) Seeds. *Archives Current Research International*, 11 (2): 1-7.
- [24] Farinde, E., Olanipekun, O. and Olasupo, B. (2018). Nutritional composition and antinutrients. Contents of raw and processed Lima bean (*Phaseolus lunatus*). *Annals of Food Science and Technology*, 19 (2): 250-264.
- [25] Alayande, L.B., Mustapha, K.B., Dabak, J. and Ubom, G.A. (2012). Comparison of nutritional values of brown and white beans in Jos North Local Government markets. *African Journal of Biotechnology*, 11 (43): 10135-10140.
- [26] Faldu, P.R., Gandhi, K.D., Patel, S. and Patel, K.G. (2017). Effect of different cooking treatment on nutritional properties of chickpea (*Cicer arietinum* L.) and kidney bean (*Phaseolus vulgaris* L.). *International Journal of Applied Agricultural and Horticultural Sciences*, 8(5): 1185-1188.
- [27] Fernandes, A., Nishida, W. and Da Costa, P. (2010). Influence of soaking on the nutritional quality of common beans (*Phaseolus vulgaris* L.) cooked with or without the soaking water. *International Journal of Food Science and Technology*, (45): 2209-2218.
- [28] Audu, S. and Aremu, M. (2011). Effect of processing on chemical composition of red kidney bean (*Phaseolus vulgaris* L.) flour. *Pakistan Journal of Nutrition*, 10 (11): 1069-1075.
- [29] Lajide, L., Oseke, M.O. and Olaoye, O.O. (2008). Vitamin C, Fibre, Lignin and Mineral Contents of Some Edible Legume Seedlings. *Journal of Food Technology*, 6:237-241.

- [30] Achu, B.L., Djuikwo, R.V., Ghomsi, T.S., Maptouom, F.C., Djouhou, F.M. and Fokou, E. (2021). Physical characteristics and the effect of boiling and fermentation on the nutritional value of *Telfairia occidentalis* Seeds. *Journal of Agricultural Chemistry and Environment*, 10: 389-401.
- [31] Bamigboye, A. and Adepoju, O. (2015). Effect of processing methods on nutritive values of Ekuru from two cultivars of beans (*Vigna Unguiculata* and *Vigna Angustifoliata*). *African Journal of Biotechnology*, 4 (21): 1790-1795.
- [32] Messina, V. (2014). Nutritional and health benefits of dried beans. *American Journal of clinical Nutrition*, 100(1): 437-442.
- [33] ENV/JM/MONO. (2015). Consensus document on compositional considerations for new varieties of common bean (*Phaseolus vulgaris* L): Key food and feed nutrients, antinutrients and other constituents; OECD environment health and safety publications, p 49.
- [34] Siddhuraju, P., Vijayakumari, K. and Janardhanan, K. (1996). Chemical composition and protein quality of little known legume, velvet bean (*Mucuna pruriens* (L) DC). *Journal of Agricultural and Food Chemistry* 44: 2636-2641.
- [35] Oraka, C. and Okoye, J. (2017). Effect of heat processing treatments on the chemical composition and functional properties of Lima bean (*Phaseolus lunatus*) Flour. *American Journal of Food Science and Nutrition*, 1: 14-24.
- [36] Duru, F., Ohaegbulam, P., Chukwudi, K. and Chukwu, J. (2020). Effect of different processing methods on the chemical, functional and phytochemical characteristics of velvet beans (*mucuna pruriens*). *International Journal of Agricultural Research and Food Production*, 5: 55-73.
- [37] Idoko, A., Onyinye, A.N., Blessing, N.O., Ayomide, T.A., Philip, O.C. and Nwali, O.N. (2020). Heating effect on phytochemical and proximate contents of cooked aqueous extract of *Phaseolus vulgaris* (Kidney beans). *Universal Journal of Pharmaceutical Research*, 4(6): 35-41.
- [38] Nkundabombi, M.G., Nakimbugwe, D. and Muyonga, J.H. (2016). Effect of processing methods on nutritional, sensory, and physicochemical characteristics of biofortified bean flour. *Food Science and Nutrition* 4(6): 384-397.
- [39] Nestares, T., Barrionuevo, M., Lopez-Frias, M. and Vidal, Concepcion. (2003). Effect of different soaking solutions on nutritive utilization of minerals (calcium, phosphorus and magnesium) from cooked beans (*Phaseolus vulgaris* L) in growing rats. *Journal of Agricultural and Food Chemistry*, 5: 515-520.
- [40] Hassan, M. and El-Syiad, S. (2014). Quality of white bean seeds (*Phaseolus vulgaris* L.) As affected by different treatments. *World Journal Dairy Food Science*, 9 (1): 20-28.
- [41] Anderson, G.J. and McLaren, G.D. (2012). Iron Physiology and Pathophysiology in Humans. Humana Press/Springer: New York, NY, USA.
- [42] Grenoble, E.F. (2010). Besoins Nutritionnels, Reseau LINUT. 103 p.
- [43] FAO-WHO (2001). Human Vitamin and Mineral Requirements. Report of a Joint FAO/OMS Expert Consultation, Bangkok, 281 p.
- [44] Onibon, V.O., Abulude, F.O. and Lawal, L. (2007). Nutritional and anti-nutritional composition of some Nigeria beans. *Journal of Food Technology*, 5 (2): 120-122.
- [45] SCF (1993). Nutrient and Energy Intakes for the European Community. Reports of the Scientific Committee for Food, Thirty-First Series (SCF), European Commission, Luxembourg.
- [46] Beto, J.A. (2015). Le rôle du calcium dans le vieillissement humain. *Clinical Nutrition Research*, 4 (1): 1-8.
- [47] Akpanyung, E.O. (2005). Proximate and mineral composition of bouillon cubes produced in Nigeria. *Pakistan Journal of Nutrition*, 4:327-329.
- [48] Otitoju, G.T.O., Otitoju, O., Nwamara, J.U. and Baiyeri, S.O. (2015). Comparative Study of the Nutrient Composition of Four Varieties of Cowpea (*Vigna unguiculata*) and Their Products (Beans-Based Products). *Pakistan Journal of Nutrition*, 14 (9): 540-546.
- [49] Farinde, E.O., Obatolu, V.A. and Fasoyiro, S.B. (2017). Microbial, Nutritional and Sensory Qualities of Baked Cooked and Steamed Cooked Lima Beans. *American Journal of Food Science Technology*, 5 (4): 156-161.
- [50] Dresbach, R.H. (1980). Handbook of poisoning, prevention, diagnosis and treatment. Dose and maximum non-effective dose based on diarrhea. *Journal of Clinical and Biochemical Nutrition*, 10: 135-44.
- [51] Chai, W. and Liebman, M. (2004). Assessment of oxalate absorption from almonds and black beans with and without the use of an extrinsic label. *Journal of Urology* 172: 953-957.
- [52] Silva, M.O., Brigide, P., Viva de Toledo, N.M., Canniatti-Brazaca, S.G. (2018). Phenolic compounds and antioxidant activity of two bean cultivars (*Phaseolus vulgaris* L.) submitted to cooking. *Brazilian Journal of Food Technology Campinas*, 21: e2016072.
- [53] Mugabo, E., Rwubatshe, B., Afoakwa, E.O. and Annor, G. (2017). Effect of pretreatments and processing conditions on anti-nutritional factors in climbing bean flours. *International Journal of Food Studies*, 6: 34-43.
- [54] Toledo NM, Rocha LC, Silva AG, Canniatti-Brazaca SGC (2013). Interaction and digestibility of phaseolin/polyphenol in the common bean. *Food Chemistry*, 138 (2-3): 776-780.
- [55] Fabbri, A.D. and Crosby, G.A. (2016). A review of the impact of preparation and cooking on the nutritional quality of vegetables and legumes. *International Journal of Gastronomy and Food Science*, 3: 2-11.
- [56] Fekadu, G.H. (2020). Nutritional and antinutritional evaluation of complementary foods formulated from maize, pea, and anchote flours. *Food Science and Nutrition*, 8: 2156-2164.
- [57] Woldegiorgis, A.Z., Abate, D., Haki, G.D. and Ziegler, G.R. (2015). Major, minor and toxic minerals and anti-nutrients composition in edible mushrooms collected from Ethiopia. *Journal of Food Processing and Technology*, 6(3): 234-244.
- [58] Olanipekun, O., Omenna, E., Olapade, O., Suleiman, P. and Omodara, O. (2015). Effect of boiling and roasting on the nutrient composition of kidney beans seed flour. *Sky Journal of Food Science* 4(2): 024-029.
- [59] Achu, B.L., Madah, S.J. and Fokou, E. (2016). Antioxidant capacity and mineral contents of five species of cucurbitaceae seeds from Cameroon. *International Journal of Recent Scientific Research*, 7(5): 10961-10970.

